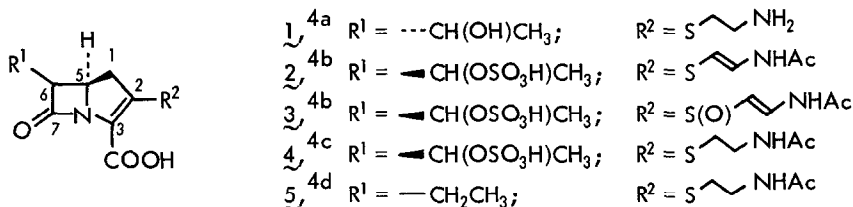


SYNTHETIC STUDIES ON β -LACTAM ANTIBIOTICS. PART 16.¹
 SYNTHESIS OF 1-CARBA-2-PENEM-3-CARBOXYLIC ACID ESTERS FROM PENICILLINS
 UTILIZING A CARBON-CARBON COUPLING REACTION

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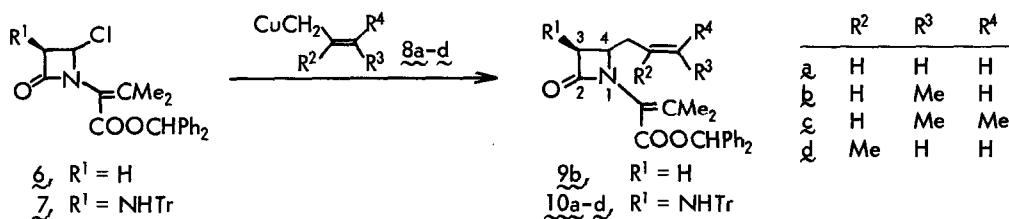
Summary: Allylazetidinones 9, 10, prepared by coupling of allylcoppers 8 with chloroazetidinones 6, 7, were converted into carbapenem esters 16, 28-31 using an Emmons-Horner reaction to introduce the 6-side chain and an intramolecular Wittig reaction to form the carbapenem ring system.

Recently, interest in the β -lactam chemistry has been focused on the syntheses^{2,3} of the carbapenem ring system, the fundamental skeleton of thienamycin 1 and its analogues⁴ 2-4. In this communication, we wish to describe a synthesis of some carbapenem esters 16, 28-31 involving a new coupling reaction of allylcoppers with chloroazetidinones derived from penicillins as a key reaction.

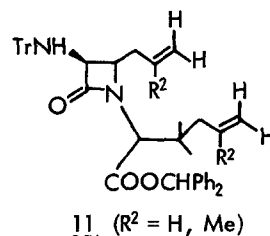


The chloroazetidinone 6⁵ or 7⁷ prepared from penicillins was allowed to react (THF, -35°C, 1-4 h then warmed to 0°C in 1 h) with allylcoppers 8a-d, prepared by mixing CuI(n-Bu₂S)₂ (ether, -76°C, N₂) with allyllithiums,⁹ giving 9b or 10a-d respectively as shown in Table I. It is noteworthy that the coupling of either isomer of 7 (4 β or 4 α) with 8d yielded a mixture of 10d with the same isomer ratio (runs 5, 6). When lithium diallylcuprates were used in place of allylcoppers, conjugated addition products 11 were formed as the major products.

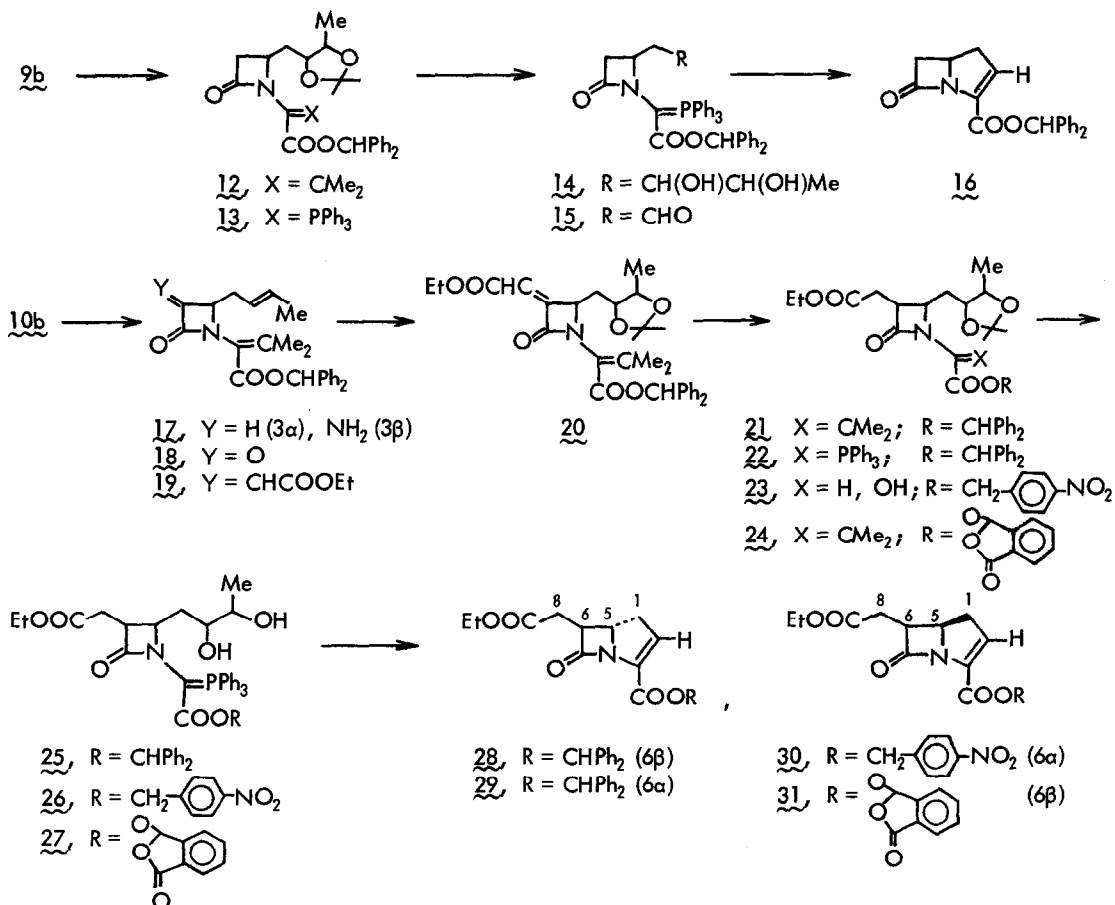
Epoxidation (m-chloroperbenzoic acid, rt) of 9b followed by hydrolysis (20% HClO₄-acetone, 0°C) and subsequent acetonidation of the resulting glycol proceeded smoothly to yield 12 (62%). Conversion of 12 into ylide 13 (70%) was performed by the method¹⁰ developed in our laboratories

Table I. Coupling of Chloride $\underline{6}$, $\underline{7}$ with Allylcopper $\underline{8a-d}$

run	chloride	allylcopper	product	yield (%)	isomer ratio ^a
1	$\underline{6}$	$\underline{8b}$	$\underline{9b}$	55.0	-
2	$4\beta\text{-}\underline{7}$	$\underline{8a}$	$\underline{10a}$	12.4	33/67
3	$4\beta\text{-}\underline{7}$	$\underline{8b}$	$\underline{10b}$	66.1	45/55
4	$4\beta\text{-}\underline{7}$	$\underline{8c}$	$\underline{10c}$	64.0	55/45
5	$4\beta\text{-}\underline{7}$	$\underline{8d}$	$\underline{10d}$	63.6	33/67
6	$4\alpha\text{-}\underline{7}$	$\underline{8d}$	$\underline{10d}$	48.2	33/67



^a The ratio of 4β - to 4α -isomers was determined by nmr.



The acetonide group of 13 was deprotected by exposure to 2 N HCl in ethanol (rt, 16 h) to give 14. Glycol fission (HIO_4 , H_2O -THF, rt, 0.5 h) of the latter gave aldehyde 15, which on neutralization (aq NaHCO_3 -AcOEt) spontaneously cyclized, giving carbapenem ester 16¹¹ (51%).

Considering that introduction of some two carbon moiety at C_6 might affect antibacterial activity, we have undertaken synthesis of C_6 -ethoxycarbonylmethylcarbapenems. The mixture of 4 β - and 4 α -10b was detritylated (p-TsOH· H_2O , acetone) and the desired product 4 β -17 (31%) was separated by silica gel chromatography from 4 α -17¹² (45%). These compounds were converted¹³ into 4 β -18 and 4 α -18, respectively (70% each), by oxidation with 3,5-di-*tert*-butyl-1,2-benzoquinone (THF, 0°C, 18 h) and subsequent hydrolysis (oxalic acid- H_2O -THF, 0°C, 21 h).

First, a model experiment using 4 α -18 of the unnatural configuration was carried out. Reaction of 4 α -18 with lithium salt of triethyl phosphonoacetate anion (THF, -20°C, 15 min) and subsequent separation of the products by silica gel chromatography yielded Z-4 α -19 (30%) and E-4 α -19 (45%), the geometries being assigned based on the chemical shifts (CDCl_3) of relevant proton signals at δ 5.83 and δ 6.48, respectively. Conversions of the butenyl groups of both Z- and E-isomers into the corresponding acetonides proceeded similarly to the case of 9b, giving Z-4 α -20 (74%) and E-4 α -20 (86%), respectively. While selective hydrogenation¹⁴ of E-4 α -20 (H_2 , 5% Pd on CaCO_3 , AcOEt, then back-esterification with Ph_2CN_2) yielded an inseparable mixture (5:1) of 3 β ,4 α - and 3 α ,4 α -21 (97%), that of Z-4 α -20 exclusively gave 3 α ,4 α -21 (87%). The mixture of 3 β ,4 α - and 3 α ,4 α -21 was converted¹⁰ into the corresponding ylides (86%), from which 3 β ,4 α -ylide 22 was separated by silica gel chromatography. The deprotection of the acetonide group of 3 β ,4 α -22 was carried out as described above giving 3 β ,4 α -25 (95%). Similarly, 3 α ,4 α -25 was obtained (78%) from 3 α ,4 α -21 via 3 α ,4 α -22. Periodic acid oxidation of 3 β ,4 α - and 3 α ,4 α -25 and subsequent neutralization of the resulting aldehydes produced carbapenem esters having the unnatural configuration, 28¹⁶ (60%) and 29 (64%),¹⁷ respectively. The stereochemistry of the C_6 -side chain of both carbapenem esters was determined by comparison of their H and ¹³C nmr spectra.

After several unsuccessful attempts to remove reductively the benzhydryl groups of esters 16, 28, and 29 and even the p-nitrobenzyl group of 30, prepared similarly from 4 β -18 via 3 α ,4 β -23, we tried to synthesize a carbapenem phthalidyl ester, which might be easily hydrolyzed by action of esterases existing in blood. Thus, carbapenem 31 was prepared from 4 β -18 via 3 β ,4 β -24 by the sequence of reactions used for converting 3 α ,4 α -21 into 29. The phthalidyl ester 31 exhibited only moderate *in vitro* antibacterial activity¹⁸ with horse blood serum, although interesting antibacterial activity of sodium C_2 -unsubstituted carbapenem carboxylates has been reported recently.^{3a,3b}

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 11. 16. IR ν (CHCl₃) 1780, 1725 cm⁻¹. NMR (T-60, CDCl₃) δ 2.67-3.0 (2H, m, C₁-H); 2.92 (1H, dd, J = 3 and 16 Hz, C_{6 α} -H); 3.47 (1H, dd, J = 6 and 16 Hz, C_{6 β} -H); 4.22 (1H, m, C₅-H); 6.50 (1H, t, J = 3 Hz, C₂-H).
 12. The trans configuration was assigned based on the nmr spectrum of its N-phenylacetyl derivative: NMR (CDCl₃) δ 3.82 (1H, dt, J = 2 and 6 Hz, C₃-H); 4.65 (1H, dd, J = 2 and 8 Hz, C₄-H).
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 16. 28. IR ν (CHCl₃) 1775, 1720 cm⁻¹. NMR (100 M/C, CDCl₃) δ 2.55-3.03 (4H, m, C₁-H, C₈-H); 3.50 (1H, ddd, J = 3, 5, and 10 Hz, C₆-H); 4.04 (1H, dt, J = 3 and 9 Hz, C₅-H); 6.47 (1H, t, J = 3 Hz, C₂-H). C¹³ NMR (CDCl₃) δ 33.2 (C₁), 35.9 (C₈), 55.2 (C₆), 58.3 (C₅).
 17. 29. IR ν (CHCl₃) 1780, 1720 cm⁻¹. NMR (100 M/C, CDCl₃) δ 2.42-2.95 (4H, m, C₁-H, C₈-H); 3.99 (1H, m, C₆-H); 4.44 (1H, dt, J = 6 and 9.5 Hz, C₅-H); 6.53 (1H, t, J = 3 Hz, C₂-H). C¹³ NMR (CDCl₃) δ 29.9 (C₁), 31.3 (C₈), 48.6 (C₆), 55.5 (C₅). $[\alpha]_D^{25}$ -38.8° (EtOH). CD (θ) (EtOH) 266 nm (-7730).
 18. The assay for antibacterial activity of the ester was carried out through the courtesy of Dr. T. Yoshida of this laboratory.

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